

RESEARCH ARTICLE

Judicious elevation of ambient carbon dioxide during hypobaric hypoxia to improve oxygenation in airline passengers: a randomized feasibility study

 Titiaan E. Post,^{1,2*} Riccardo De Gioannis,^{1,3*}  Daniel Rooney,¹  Martin Wittkowski,¹  Patrick Lau,¹ Leopold Lecheler,¹  Jens Jordan,^{1,4}  Jochen Zange,^{1,4}  Jörn Rittweger,^{1,4†} and  Daniel Aeschbach^{1,5}

¹Department of Sleep and Human Factors Research, Institute of Aerospace Medicine, German Aerospace Center (DLR), Cologne, Germany; ²Centre for Human Drug Research (CHDR), Leiden, The Netherlands; ³Department III for Internal Medicine, Heart Center, University Hospital Cologne, Cologne, Germany; ⁴Medical Faculty, University of Cologne, Cologne, Germany; and ⁵Institute of Experimental Epileptology and Cognition Research, University of Bonn Medical Center, Bonn, Germany

Abstract

Despite pressurization of airliner cabins, some passengers experience in-flight hypobaric hypoxia with blood oxygen saturation dropping below 90%, potentially causing discomfort and increasing the risk of medical events. Enrichment of the cabin air with carbon dioxide (CO₂) may augment passengers' blood and tissue oxygenation by stimulating respiratory drive, thereby increasing health and safety during air travel. In a randomized double-blind crossover study, we exposed 17 healthy adults (8 women; age range: 18–40 yr) on separate days to two ambient CO₂ levels (0.1% vs. 1.0% indoor sea-level equivalents; 0.76 vs. 7.60 mmHg partial pressure) during 6 h of hypobaric hypoxia (~565 mmHg total barometric pressure; corresponding to 2,438 m altitude) in an altitude chamber simulating long-haul flight conditions. We measured oxygen saturation of the blood (SpO₂), brain, and muscle (tissue saturation index derived from near-infrared spectroscopy), respiration, and cognitive function (sustained attention, working memory, and hand-eye coordination) hourly. In addition, we conducted capillary blood gas analyses at baseline, 15 min, and 6 h after hypoxia onset. During hypobaric hypoxia, ambient CO₂ enrichment on average increased Pco₂ from 36.3 to 38.3 mmHg, Po₂ from 60.9 to 68.3 mmHg, and minute ventilation from 9.7 to 10.4 L/min, whereas reducing the time fraction of SpO₂ < 90% from 18.8% to 2.5%. Tissue oxygenation increased in the brain from 62.8% to 63.9%, whereas no change was found in the muscle. High ambient CO₂ had no effect on cognitive performance. Taken together, enrichment of cabin air with CO₂ during hypobaric hypoxia may improve blood and brain oxygenation.

NEW & NOTEWORTHY Elevating ambient CO₂ can improve blood and brain oxygenation in healthy individuals during moderate hypoxia as experienced in an airliner cabin. The findings offer a new approach potentially benefiting the health and safety of airline passengers and support investigating CO₂ enrichment in broader passenger populations, especially in those at risk of hypoxemia. Exploration of CO₂-based interventions to enhance passenger oxygenation may also prompt regulatory reassessment of cabin CO₂ limits and technological innovations in ventilation systems.

aircraft cabin pressure; ambient CO₂; blood oxygen saturation; hypobaric hypoxia; respiratory rate

INTRODUCTION

Commercial jet aircraft often fly at altitudes of 30,000–43,000 ft. At such altitudes, oxygen partial pressure outside the airliner cabin is profoundly reduced and incompatible with human life. Therefore, aircraft cabins are pressurized with the corresponding maximum cabin altitude during normal operations, not to exceed 8,000 ft (~2,438 m) (1, 2). Cabin pressurization ensures adequate oxygenation for most passengers, with oxygen saturation (SpO₂) under these conditions typically ranging between 91% and 95%, which is

considered safe. Most guidelines define hypoxemia as an oxygen saturation level below 90% that should be avoided (3). However, existing atmospheric conditions in the cabin fail to prevent hypoxemia in all passengers effectively. In some individuals, especially in those with respiratory or cardiopulmonary conditions, oxygen saturation can decrease below 90% (4–7). Moreover, during sleep under flight conditions, blood oxygenation can drop below 90% for a sizable fraction of time, even in healthy individuals (4, 7, 8). Hypoxemia can exacerbate medical conditions or incite medical events (9, 10). Increased mortality risk has been observed in emergency



†Deceased April 2025.

*T. E. Post and R. De Gioannis contributed equally to this work.

Correspondence: D. Aeschbach (daniel.aeschbach@dlr.de).

Submitted 19 December 2025 / Revised 5 June 2026 / Accepted 15 June 2026



admissions with SpO₂ levels between 86% and 89% compared with 90% or higher (10), and postoperative outcomes worsen when desaturation occurs below 90% (11). Moreover, cardiac and respiratory complaints are among the most serious in-flight medical events (9).

One potential solution for reducing the occurrence of in-flight hypoxemia is to increase cabin pressure. Indeed, most airliners nowadays operate at a lower cabin altitude than is maximally allowed (12). However, a further increase in cabin pressure is limited, as this measure would increase aircraft weight and fuel consumption due to the need for a reinforced cabin structure, subsequently worsening its environmental impact (12). Given the continuously increasing air travel volume, alternative strategies to improve oxygenation in passengers are currently being explored.

We assessed the potential of enriching the cabin atmosphere with carbon dioxide (CO₂) as a countermeasure to in-flight hypoxia. Our objective was to assess whether an increase in ambient CO₂ improves blood SpO₂ in healthy individuals during hypobaric hypoxia, as typically experienced during air travel (1). Exposure to reduced oxygen partial pressure (PO₂) elicits an increase in ventilation, the so-called “hypoxic ventilatory response.” The ventilatory response increases CO₂ elimination, leading to hypocapnia, which in turn opposes respiratory drive. Consequently, in most people exposed to modest hypoxia, ventilation stabilizes slightly above baseline levels (13). Although these counteracting regulatory circuits maintain overall homeostasis, they can limit the ability to cope with hypoxia. Conversely, increased inspired CO₂ prevents hypocapnia and augments ventilation under hypoxia for prolonged periods of time (14). Moreover, because CO₂ is a potent cerebral vasodilator, maintaining adequate levels of arterial CO₂ can help ensure proper brain perfusion and prevent light-headedness, thereby supporting overall brain function and alertness (15).

Our primary hypothesis was that during a realistic simulation of a 6-h flight, a sea-level equivalent of 1% CO₂ inside the airliner cabin increases our primary outcome, i.e., blood SpO₂, in healthy individuals compared with a flight condition without CO₂ enrichment (control). To gain insight into the underlying mechanisms, we measured secondary endpoints, including capillary blood gases and respiration. In addition, we estimated brain and skeletal muscle oxygen saturation using near-infrared spectroscopy (NIRS). Finally, we evaluated cognitive performance as reflected in sustained attention, working memory, and hand-eye coordination.

MATERIALS AND METHODS

We obtained informed written consent from study participants before the start of the study. The protocol was approved by the ethics committee of the North Rhine Medical Association (Ärztzammer Nordrhein; Protocol No.: 2018253) and prospectively registered at the German Clinical Trials Register (DRKS, Registration No.: DRKS00015820). We conducted all procedures according to the Declaration of Helsinki (2013). Study participants were compensated for their participation. We performed the study as well as the medical screening at the Institute of Aerospace Medicine, German Aerospace Center (DLR) in Cologne, Germany.

Participants

Men and women aged 18–40 yr with a body mass index (BMI) between 18 and 30 kg/m² were eligible for the study. Participants were enrolled according to a predefined set of inclusion and exclusion criteria. In brief, participants were in good health, as established by a physical examination, medical history, stress-electrocardiogram, and routine blood and urine testing. Participants reported no recent exposure to altitudes above 1,500 m within 6 wk before the study, no regular practice of high-performance sports, and did not play wind instruments. Participants abstained from alcohol, nicotine, drugs, and caffeine for 1 wk before each visit to the laboratory, which was verified by urine testing upon admission. Here we report on one of the two study arms (~8,000 ft condition, see Study Design), which included 17 participants (8 women, mean age ± SD: 27.8 ± 4.2 yr, mean BMI ± SD: 24.6 ± 3.2).

Study Design

Participants were randomized to one of two study arms—one simulating a cabin pressure equivalent to the air pressure at an altitude of ~8,000 ft and the second arm simulating a pressure equivalent to ~14,000 ft (Fig. 1). In each study arm, participants underwent two ambient CO₂ conditions (0.76 and 7.60 mmHg) on separate days with a 7-day wash-out period, according to a randomized, double-blind, cross-over design. Due to the occurrence of adverse events, the second study arm (~14,000 ft) was discontinued after 7 participants (see RESULTS). During both conditions of the first study arm, participants spent 6 h in an altitude chamber, which was depressurized to simulate an altitude of ~8,000 ft (565 mmHg), corresponding to the maximum cabin altitude of an airliner as allowed during normal operation. The two conditions differed by the ambient CO₂ concentration in the altitude chamber, consisting of either a low or high CO₂ condition with normalized indoor sea-level (760 mmHg) equivalents of 0.1% (0.76 mmHg) or 1.0% (7.60 mmHg) CO₂. We randomly assigned participants to the two sequences in which the high and low CO₂ conditions occurred. Participants were initially grouped (groups of 2) based on their availability. Subsequently, the order of treatment conditions was randomized across these groups using an R script that simulates a coin toss. Both the physician taking the measurements and the participants inside the chamber were blinded to the treatment conditions. In contrast, the study staff operating the chamber from outside were unblinded.

We collected baseline data every morning on each study day under normobaric and normoxic conditions. The experiment in the pressure chamber was divided into six test blocks, each lasting 60 min. A test block comprised the Environmental Symptoms Questionnaire (ESQ) and the 0–10 Borg CR10 Scale (3 min), a cognitive test battery (22 min), physiological measurements (20 min), and a break (15 min). Physiological measurements included spirometry, pulse oximetry, and near-infrared spectroscopy (NIRS) that were recorded simultaneously. During each break, participants received hourly isocaloric snacks and had the opportunity to use the restroom within the altitude chamber. In addition, we performed capillary blood gas analysis (cBGA) 15 min and 6 h after the start of hypoxia exposure.

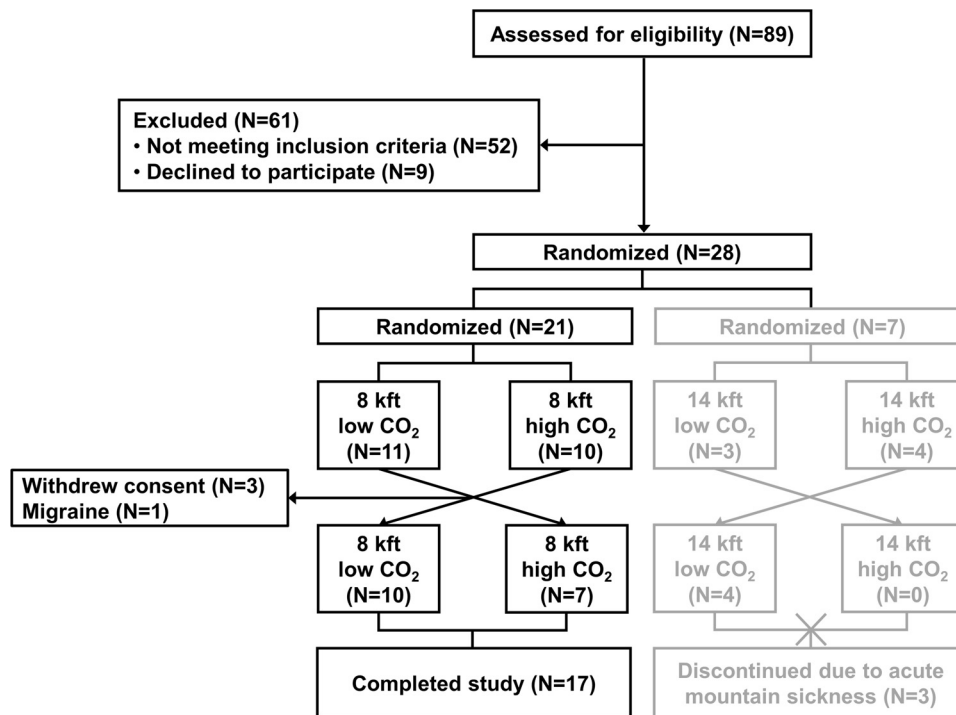


Figure 1. Flow diagram of the study.

Capillary Blood Gas Analyses

We preferably collected capillary blood from the earlobe 5 min after application of a blood flow-stimulating ointment (Finalgon, Sanofi-Aventis, Frankfurt, Germany). In rare cases, we used the finger tip as an alternative sampling site. We analyzed samples using the epoc Blood Analysis System (Siemens Healthineers, Erlangen, Germany), which involves electrodes to assess the concentration of gases in the blood.

Spirometry and Pulse Oximetry

We measured ventilation and gas-exchange parameters (O₂ and CO₂) through an oral-nasal mask using a breath-by-breath spirometer (Inn00400, Innovision, Glamsbjerg, Denmark). In between hourly spirometry recordings, the participants did not wear the mask. After conducting our analysis, we discovered that the spirometer encountered problems when exposed to high ambient CO₂ levels (7.60 mmHg) under hypobaric conditions. After consulting with the manufacturer, we chose to exclude all data that depended on CO₂ measurements of the Innovision device. In the absence of end-expiratory CO₂ measurements, we were unable to calculate alveolar ventilation. Therefore, we focused on minute ventilation as a key mediator in optimizing oxygen saturation. We assessed blood oxygen saturation (SpO₂) with a finger pulse oximeter (Nonin, Plymouth, MN).

Near-Infrared Spectroscopy

We placed sensors (Portalite and PortaMon, Artinis Medical Systems, Elst, the Netherlands) over the right frontal eminence of the skull and the belly of the right vastus lateralis muscle. We securely fixed both sensors with dark elastic tape to minimize artifacts from external light. The devices measured the tissue concentration of oxygenated (HbO₂, μmol/L

tissue) and deoxygenated hemoglobin (Hb, μmol/L tissue) at a sampling rate of 10 Hz. From these values, total hemoglobin (tHb = HbO₂ + Hb, μmol/L tissue) and tissue oxygen saturation index (TSI; 100 × HbO₂/tHb) were determined.

Cognitive Performance

The cognitive test battery used in this study comprised three components: a 10-min version of the psychomotor vigilance task (PVT; 16, 17) to measure sustained attention, the N-back task with varying difficulty levels (1-, 2-, and 3-back load) to evaluate working memory (18, 19), and the adaptive tracking task to assess hand-eye coordination (20, 21). The selected measures of interest included PVT speed (s⁻¹), PVT number of lapses (response time > 500 ms), 2-back number of omissions, and the mean score obtained from the adaptive tracking task. To mitigate the potential impact of training effects, each task was performed three times before the study.

Statistical Analysis

A statistical power analysis for the first arm was performed assuming a moderate effect of $f = 0.18$ for the effect of ambient CO₂ on blood oxygen saturation during hypoxia exposure (high vs. low ambient CO₂). The analysis was based on a repeated measures analysis of variance (ANOVA) with $\alpha = 0.05$ and power = 0.80. The projected sample size needed was approximately $n = 17$ for the crossover design in each arm. We analyzed the data using R, version 4.0.1 (R Foundation, Vienna, Austria). Numerical data are presented as means ± standard errors of the means unless indicated otherwise. Repeatedly measured data were analyzed with linear mixed-effects models with treatment, time, and treatment-by-time as fixed effects, and participant-specific intercepts. Measurements immediately preceding the hypoxia exposure served as baseline and

were used as covariates in the models. In cases where the interaction term was not significant, a simplified model excluding the interaction term is reported. Post hoc comparisons were done on estimated marginal means. Data shown in the figures represent descriptive means. Normality was assessed using residual plots. An alpha value of 0.05 was considered significant, with correction for multiplicity performed using the Bonferroni–Holm method. The first 5 min of physiological recordings in each test block were excluded from the analysis to minimize artifacts due to the application of the measurement equipment. The number of PVT lapses and 2-back omissions resulted in a considerable number of zeros. To account for the excess of zeros in the count variable PVT lapses, we used the zero-inflated negative binomial (ZINB) model. For the 2-back omissions, the ZINB model returned no overdispersion, and therefore a zero-inflated Poisson model was applied.

Missing Data

Two participants had missing SpO₂ data in a total of five test blocks and one baseline measurement due to a sensor connection failure with the spirometry device. As the average intra-individual baseline differences were negligible (0.1% SpO₂), we imputed the missing baseline value using the available baseline data from the same individual on the other experimental day. In addition, for a third participant, all spirometry and SpO₂ data from one test block were missing due to device failure. Overall, the missing data accounted for 3% of our dataset, and their occurrence could reasonably be considered random.

RESULTS

Between December 2018 and July 2019, 89 interested volunteers entered a multistep screening procedure, resulting in the selection of 28 healthy adults. The study was initially designed with two arms—one simulating a cabin pressure equivalent to the air pressure at an altitude of ~8,000 ft and the second arm simulating a pressure equivalent to ~14,000 ft (Fig. 1). However, due to signs of acute mountain sickness in three participants, we discontinued exposure to 14,000 ft after testing seven persons. Consequently, data collected from this arm were excluded from subsequent analyses. As a result, twenty-one participants were assigned to the 8,000 ft arm. Within this group, four participants did not continue the study after the first visit (low CO₂ condition in each case). Of those, three participants withdrew their consent, whereas one experienced migraine and was excluded. Consequently, the final dataset for the 8,000 ft arm included 17 complete datasets (8 women, mean age ± SD: 27.8 ± 4.2 yr, mean BMI ± SD: 24.6 ± 3.2).

In hypoxia, SpO₂ values were higher in high compared with low ambient CO₂ (Fig. 2A, mixed model, $P < 0.001$). Time fraction of SpO₂ < 90% was lower in high compared with low ambient CO₂ (Fig. 2B, paired t test, $P = 0.005$). High ambient CO₂ resulted in increased capillary blood levels of PCO₂ and PO₂ compared with low ambient CO₂ (Fig. 2, C and D, mixed model, factor treatment $P < 0.001$ in both cases), accompanied by a decrease in pH levels (Fig. 2E, mixed model, $P = 0.006$), and a tendency toward increased bicarbonate levels (Fig. 2F, mixed model, $P = 0.077$). There

were no significant interactions between treatment and time for any variable.

Although the present study was not designed and powered to detect potential sex differences, we carried out explorative analyses by including sex as a covariate in the mixed-model analyses of SpO₂, PO₂, and PCO₂. No sex differences were observed except for a tendency toward lower PCO₂ in females than in males (34.1 vs. 38.6 mmHg, $P = 0.051$). There were no significant interactions between treatment and sex.

Changes from baseline in minute ventilation, respiratory rate, and tidal volume are shown in Fig. 3. High ambient CO₂ levels resulted in increased minute ventilation and respiratory rate (factor treatment $P < 0.001$ in both cases), along with a tendency toward decreased tidal volume (factor treatment $P = 0.061$). There were no significant interactions between treatment and time for any variable.

Individual trajectories between ambient CO₂ conditions are shown for SpO₂ and PCO₂ (Fig. 4A) and for SpO₂ and minute ventilation (Fig. 4B). Fifteen out of 17 participants (88%) showed an increase in SpO₂ with increasing PCO₂, and 13 participants (76%) showed an increase in SpO₂ with increasing minute ventilation. Corresponding trajectories for PO₂ were somewhat less tight: 12 participants (71%) showed an increase in PO₂ with increasing PCO₂, and 11 participants (65%) showed an increase in PO₂ with increasing minute ventilation (data not shown).

High ambient CO₂ resulted in higher brain tissue saturation index (TSI; Fig. 5A, factor treatment $P = 0.003$), with the largest difference occurring 2 h after hypoxia onset. Muscle TSI showed no difference between treatments (Fig. 5B). In both brain and muscle, TSI varied over time (factor time $P < 0.001$ in both cases). There were no significant interactions between treatment and time in either tissue. We observed no significant differences for tHb and HbO₂.

Cognitive performance measurements did not differ significantly between low and high ambient CO₂ (Supplemental Fig. S1). The time fraction of SpO₂ < 90% did not significantly correlate with cognitive performance measures.

No significant differences in the acute mountain sickness–cerebral score derived from the ESQ were observed between treatments (Supplemental Fig. S2). In high CO₂, two participants reported slight breathlessness (Borg score = 2), whereas none did so in the low CO₂ conditions. No participant reported symptoms above slight breathlessness in any condition.

DISCUSSION

CO₂ enrichment of the ambient air during moderate hypobaric hypoxia, resembling conditions in an airliner cabin, improved blood oxygenation. The intervention was well tolerated in healthy young individuals. High CO₂ elevated minute ventilation and respiratory rate. Consequently, CO₂ enrichment increased blood gas levels of PCO₂ and PO₂, as well as SpO₂, while reducing the time fraction of SpO₂ < 90%. Furthermore, CO₂ increased tissue oxygenation in the brain but not in skeletal muscle. There was no cognitive impairment in both conditions compared with normoxia.

Previous work has shown a rise in tidal volume with moderate increases in ambient CO₂, whereas in the current study, changes in ventilation were attributed to an increase in

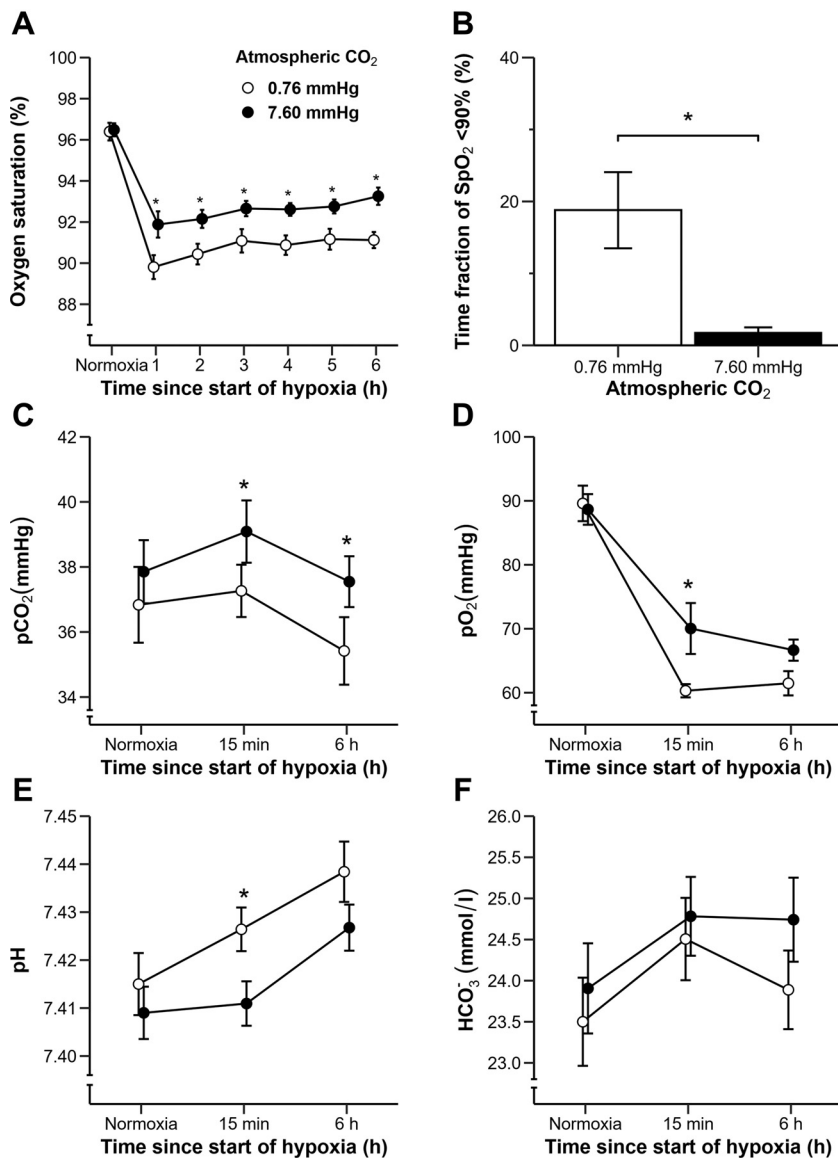


Figure 2. Blood oxygenation and blood gases during 6 h of hypobaric hypoxia with low and high ambient CO₂. **A:** mean ($\pm$ SE, $n = 17$ participants) blood oxygen saturation (SpO₂) at hourly intervals during exposures. **B:** time fraction of SpO₂ < 90% during exposures calculated as % of the total time of SpO₂ measurements. Carbon dioxide partial pressure (C), oxygen partial pressure (D), pH (E), and bicarbonate (F) levels in capillary blood at baseline, after 15 min, and 6 h of exposure. *Significant differences between conditions ($P < 0.05$, marginal means contrasts with Bonferroni-Holm correction except for B where a paired t test was used) between conditions post Bonferroni-Holm correction. For results of mixed-model analysis, see RESULTS.

respiratory rate only. It is possible that hypobaric hypoxia, conceivably due to the reduced airway resistance to less dense air, contributed to a shift in the balance between depth and frequency of breathing with higher CO₂. Moreover, subliminal discomfort/anxiety associated with the instrumentation and the unusual and confined environment of the altitude chamber may have reinforced a bias toward faster and shallower breathing with increased CO₂.

In addition to stimulating ventilation, inspired CO₂ may have improved pulmonary gas exchange by enhancing ventilation-perfusion ($\dot{V}_A/\dot{Q}$) matching. By influencing pulmonary vascular tone and augmenting hypoxic pulmonary vasoconstriction, CO₂ may have redistributed blood flow toward better-ventilated alveoli, reducing $\dot{V}_A/\dot{Q}$ mismatch and enhancing overall oxygenation. Furthermore, CO₂-induced bronchodilation may also contribute to improving $\dot{V}_A/\dot{Q}$ matching (22).

Exposure of unacclimatized persons to simulated moderate altitude has been associated with reduced SpO₂ and increased frequency of symptoms, including altitude-related

malaise, muscular discomfort, and fatigue, which became manifest after 3–9 h of exposure (6). The slight increase in blood and brain oxygenation during CO₂ enrichment observed in our study may contribute to mitigating some of these effects. Although the study by Muhm et al. (6) as well as the current study used SpO₂ as the primary oxygen marker, it should be noted that SpO₂ is not always reliable when determining whether an individual is hypoxic or not (23). PaO₂, which is important for the delivery through diffusion of oxygen from blood to the cells, can deviate considerably from the levels of SpO₂ during hypoxic events.

It is tempting to speculate that, beyond its role in improving oxygenation under hypobaric hypoxia, controlled enrichment of emergency breathing air with CO₂ could have practical applications during in-flight decompression events. By sustaining respiratory drive, CO₂ may improve the effectiveness of emergency oxygen delivery, particularly during depressurization at very high altitudes. This intervention could extend the safe duration for passengers and crew to tolerate hypobaric conditions in situations where emergency

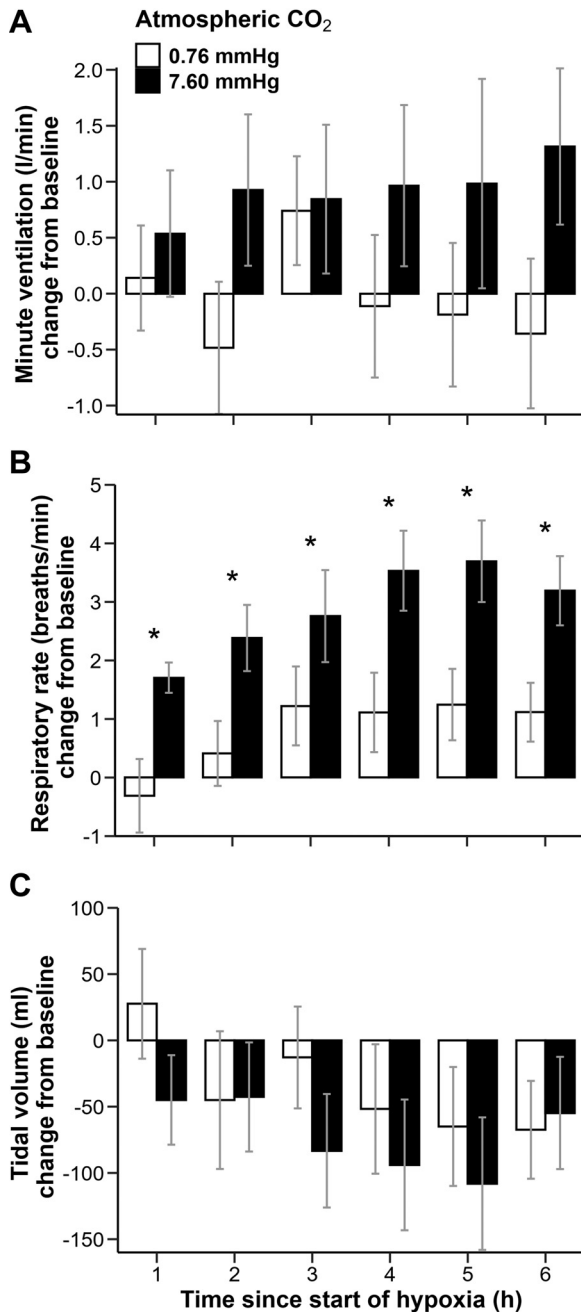


Figure 3. Changes in ventilation during 6 h of hypobaric hypoxia with low and high ambient CO₂. Data show mean ($\pm$ SE, $n = 17$ participants) changes from baseline (normoxia). Minute ventilation (calculated as the product of tidal volume and respiratory rate) in L/min (A), respiratory rate in breaths per minute (B), and tidal volume in mL (C). *Significant differences between conditions ($P < 0.05$, marginal means contrasts with Bonferroni-Holm correction) between conditions post Bonferroni-Holm correction. For results of mixed-model analysis, see RESULTS.

descent is constrained by high terrain, potentially enabling aircraft to operate over high-terrain regions—such as the Himalayas—that are otherwise restricted due to altitude-related safety concerns.

The present study indicates that the physiological impact of ambient CO₂ could be leveraged to maintain oxygen saturation levels of airline passengers above 90%. Although acknowledging potential adverse effects of ambient CO₂, we recognize the

need for a nuanced discussion. Increased inhaled ambient CO₂ can lead to adverse effects (24, 25). Aviation authorities currently mandate a cabin CO₂ limit of 0.5% (26). Under hypoxic conditions and with a controlled ambient CO₂ concentration of 1%, we did not observe impairments in cognitive function or acute well-being. By providing a comprehensive perspective, our data align with the proposal to use 1% ambient CO₂ to maintain passenger well-being during flights. Nonetheless, it is important to recognize that the current investigations need to be extended to individuals with medical conditions and those of older age.

A practical implementation of the approach outlined in this paper could be increasing cabin air recirculation to leverage the CO₂ generated by passengers. On the one hand, increasing cabin air recirculation lessens the need for air sourced from the aircraft's engines (so-called bleed air) for ventilation purposes, as proposed by the European Union's Clean Sky initiative (27). Positive effects would include lower fuel consumption, increased engine thrust, and reduced risk of toxic-fume events (12, 28). On the other hand, it might lead to concerns about increased odors and the heightened risk of spreading infectious diseases due to lower air exchange rates (29). However, these challenges can be overcome with straightforward technological adaptations to the aircraft's air conditioning systems, such as the integration of CO₂ sensors and specialized air filters (30, 31).

Harvey et al. (32) proposed the inhalation of 3% CO₂ as a potential therapy for mountain sickness, reporting symptom relief and increases in PaCO₂ of 4.5 mmHg and PaO₂ of 17.2 mmHg. A follow-up study confirmed these findings, although changes were smaller with increases in PaCO₂ of 1.0 mmHg and PaO₂ of 5.7 mmHg (33). Our research aligns with these previous studies that used a similar approach of increasing ambient CO₂, further supporting the validity and effectiveness of this method. It should be kept in mind, however, that these previous studies were carried out in climbers at considerably higher altitudes.

Previous studies have reported a favorable impact of ambient CO₂ on cerebral oxygen saturation in the presence of hypoxia and normoxia (34, 35). Our NIRS measurements on brain tissue support these findings, suggesting a temporary vasodilation and/or the effect of increased respiration due to high CO₂.

During the 6-h exposure at ~8,000 ft, there were no significant impairments observed in cognitive performance when compared with baseline under normobaric normoxia. A recent review revealed only minor impairments in some cognitive domains at oxygen levels of 80%–89% SpO₂ (36). Moreover, the addition of 7.60 mmHg ambient CO₂ did not cause cognitive impairments, indicating no apparent risk to cognition in healthy individuals.

Although the current work is physiologically grounded, it draws particular strength from its application to aeronautics. Nonetheless, a limitation of our study is that we only included younger and healthy participants who abstained from alcohol, caffeine, and nicotine, which limits the generalizability of our study. Our findings cannot simply be extrapolated to older individuals and those with medical conditions. However, it is these older and diseased individuals who are at a higher risk of experiencing in-flight hypoxemia and who may benefit the most from in-flight measures

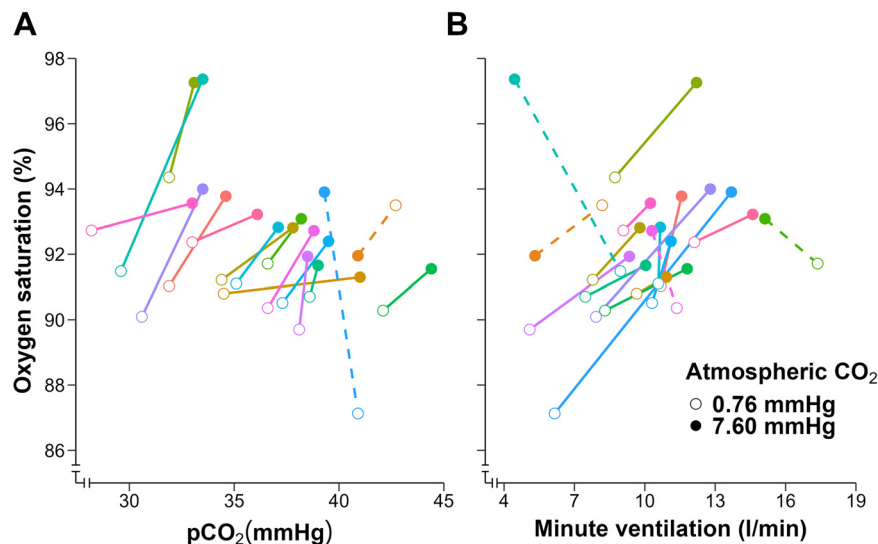


Figure 4. Individual trajectories of oxygen saturation during hypobaric hypoxia between low and high ambient CO₂. Data were obtained 6 h after hypoxia onset. Oxygen saturation plotted as a function of PCO₂ (A), and as a function of minute ventilation (B). Dashed lines highlight participants deviating from the predominant direction observed in the majority of participants (solid lines). In one participant, the final 6 min of SpO₂ recordings were removed before averaging due to artifacts/implausible data. SpO₂, oxygen saturation.

to improve oxygenation. Passengers with respiratory diseases (e.g., chronic obstructive pulmonary disease, sleep apnea) and cardiovascular diseases (e.g., cardiomyopathy), in particular, are more vulnerable to the dangers of in-flight hypoxemia, which could prevent them from flying at all (37–39). Such increased vulnerability underscores the importance of studying their capacity to tolerate elevated ambient CO₂ levels. In clinical settings, inhalation of CO₂ at concentrations of up to 3% has shown successful therapeutic

outcomes for conditions such as Cheyne–Stokes respiration and central sleep apnea (40). For passengers with respiratory diseases, the effects of increased ambient CO₂ are likely to vary depending on the underlying pathophysiology of their specific condition. Thus, it is crucial to not only evaluate the potential benefits of 1% inspiratory CO₂ in increasing oxygen saturation but also consider its potential to exacerbate the condition of these passengers. Another limitation is the absence of end-expiratory CO₂ measurements, which

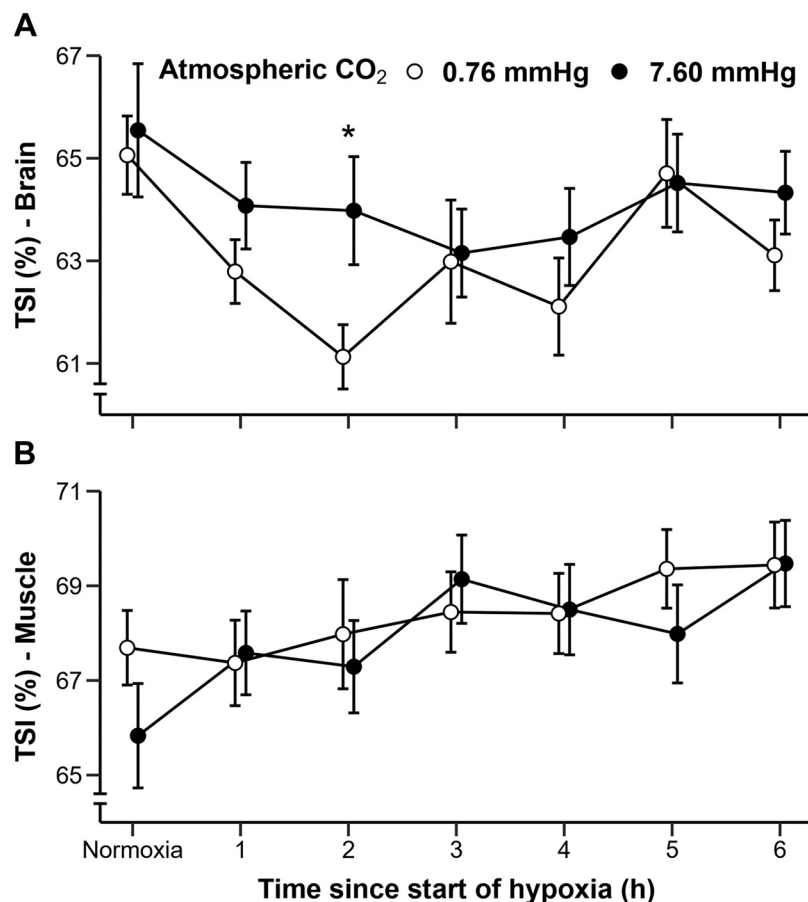


Figure 5. Tissue saturation index (TSI) derived from near-infrared spectroscopy during hypobaric hypoxia with low and high ambient CO₂. Data show means (±SE, *n* = 17 participants) for brain (A) and muscle (B). *Significant differences between conditions (*P* < 0.05, marginal means contrasts with Bonferroni–Holm correction) between conditions post Bonferroni–Holm correction. For results of mixed-model analysis, see RESULTS.

prevented us from accurately calculating alveolar ventilation in addition to minute ventilation.

Conclusions

With the popularity of air travel as a major transportation mode, atmospheric conditions within aircraft cabins should be optimized to ensure the well-being of all passengers. Our study challenges existing aviation regulations by showing that an increase of CO₂ concentration to a normalized sea-level equivalent of 1% improved mean blood oxygenation in the majority of the healthy individuals studied here. Notably, our findings demonstrate that a higher cabin CO₂ concentration effectively prevents mean blood oxygen saturation from dropping below the clinical threshold of 90%. However, considering the diverse and changing demographics of airline passengers, including more individuals who are older and/or have medical conditions, the safety implications of a 1% CO₂ concentration should be thoroughly investigated across different groups of passengers.

DATA AVAILABILITY

Source data for this study are openly available in the Open Science Framework at <https://doi.org/10.17605/OSF.IO/M8GQ9>.

SUPPLEMENTAL MATERIAL

Supplemental Figs. S1 and S2: <https://doi.org/10.17605/OSF.IO/M8GQ9>.

ACKNOWLEDGMENTS

We thank the study participants and Dr. Ulrich Limper for expert comments during the revision process. This work is written in memory of our colleague and friend, Prof. Dr. Jörn Rittweger.

GRANTS

This study was part of the Advanced Technologies Long Range Aircraft project and funded by the Aeronautics Program of the German Aerospace Center.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

T.E.P., R.D.G., D.R., M.W., P.L., L.L., J.J., J.Z., J.R., and D.A. conceived and designed research; T.E.P., R.D.G., D.R., M.W., P.L., and L.L. performed experiments; T.E.P., R.D.G., D.R., M.W., L.L., J.Z., and D.A. analyzed data; T.E.P., R.D.G., D.R., M.W., L.L., J.Z., and D.A. interpreted results of experiments; T.E.P. and D.A. prepared figures; T.E.P., R.D.G., D.R., and D.A. drafted manuscript; T.E.P., R.D.G., D.R., M.W., P.L., J.J., J.Z., J.R., and D.A. edited and revised manuscript; T.E.P., R.D.G., D.R., M.W., P.L., J.J., J.Z., J.R., and D.A. approved final version of manuscript.

REFERENCES

- Hampson NB, Kregenow DA, Mahoney AM, Kirtland SH, Horan KL, Holm JR, Gerbino AJ. Altitude exposures during commercial flight: a reappraisal. *Aviat Space Environ Med* 84: 27–31, 2013. doi:10.3357/asm.3438.2013.
- Cottrell JJ. Altitude exposures during aircraft flight. Flying higher. *Chest* 93: 81–84, 1988. doi:10.1378/chest.93.1.81.
- O'Driscoll BR, Howard LS, Earis J, Mak V, BTS Emergency Oxygen Guideline Development Group. BTS guideline for oxygen use in adults in healthcare and emergency settings. *Thorax* 72: 111–119, 2017. doi:10.1136/thoraxjnl-2016-209729.
- Elmenhorst EM, Rooney D, Benderoth S, Wittkowski M, Wenzel J, Aeschbach D. Sleep-induced hypoxia under flight conditions: implications and countermeasures for long-haul flight crews and passengers. *Nat Sci Sleep* 14: 193–205, 2022. doi:10.2147/NSS.S339196.
- Rooney D, Herkenrath S, Priegnitz C, Putzke M, Tremi M, Wenzel J, Aeschbach D, Randerath W. Choosing an adequate test to determine fitness for air travel in obese individuals. *Chest* 156: 926–932, 2019. doi:10.1016/j.chest.2019.07.022.
- Muhm JM, Rock PB, McMullin DL, Jones SP, Lu IL, Eilers KD, Space DR, McMullen A. Effect of aircraft-cabin altitude on passenger discomfort. *N Engl J Med* 357: 18–27, 2007. doi:10.1056/NEJMoa062770.
- Trammer RA, Rooney D, Benderoth S, Wittkowski M, Wenzel J, Elmenhorst E-M. Effects of moderate alcohol consumption and hypobaric hypoxia: implications for passengers' sleep, oxygen saturation and heart rate on long-haul flights. *Thorax* 79: 970–978, 2024. doi:10.1136/thorax-2023-220998.
- Post TE, De Gioannis R, Schmitz J, Wittkowski M, Schäper TM, Wrobeln A, Fandrey J, Schmitz M-T, Takahashi JS, Jordan J, Elmenhorst E-M, Aeschbach D. Resetting of the human circadian melatonin rhythm by ambient hypoxia. *J Pineal Res* 77: e70029, 2025. doi:10.1111/jpi.70029.
- Silverman D, Gendreau M. Medical issues associated with commercial flights. *Lancet* 373: 2067–2077, 2009. doi:10.1016/S0140-6736(09)60209-9.
- Goodacre S, Turner J, Nicholl J. Prediction of mortality among emergency medical admissions. *Emerg Med J* 23: 372–375, 2006. doi:10.1136/emj.2005.028522.
- Rostin P, Teja BJ, Friedrich S, Shaefi S, Murugappan KR, Ramachandran SK, Houle TT, Eikermann M. The association of early postoperative desaturation in the operating theatre with hospital discharge to a skilled nursing or long-term care facility. *Anaesthesia* 74: 457–467, 2019. doi:10.1111/anae.14517.
- Bagshaw M, Illig P. The aircraft cabin environment. In: *Travel Medicine* (4th ed.), edited by Keystone JS, Kozarsky PE, Connor BA, Nothdurft HD, Mendelson M, Leder K. Elsevier, 2019, p. 429–436.
- Bigard G, Forster H. Ventilatory responses to acute and chronic hypoxia. *Compr Physiol* 1994: 1207–1239, 2011. doi:10.1002/j.2040-4603.1994.tb01200.x.
- Howard LS, Robbins PA. Ventilatory response to 8 h of isocapnic and poikilocapnic hypoxia in humans. *J Appl Physiol* (1985) 78: 1092–1097, 1995. doi:10.1152/jappl.1995.78.3.1092.
- Rafferty GF, Saisch SGN, Gardner WN. Relation of hypocapnic symptoms to rate of fall of end-tidal Pco₂ in normal subjects. *Respir Med* 86: 335–340, 1992. doi:10.1016/s0954-6111(06)80033-8.
- Dinges DF, Powell JW. Microcomputer analyses of performance on a portable, simple visual RT task during sustained operations. *Behav Res Methods, Instrum Comput* 17: 652–655, 1985. doi:10.3758/BF03200977.
- Benderoth S, Hörmann HJ, Schießl C, Elmenhorst EM. Reliability and validity of a 3-min psychomotor vigilance task in assessing sensitivity to sleep loss and alcohol: fitness for duty in aviation and transportation. *Sleep* 44: zsab151, 2021. doi:10.1093/sleep/zbab151.
- Gevins A, Cuttito B. Spatiotemporal dynamics of component processes in human working memory. *Electroencephalogr Clin Neurophysiol* 87: 128–143, 1993. doi:10.1016/0013-4694(93)90119-g.
- Hennecke E, Lange D, Steenbergen F, Fronczek-Ponczet J, Elmenhorst D, Bauer A, Aeschbach D, Elmenhorst E-M. Adverse interaction effects of chronic and acute sleep deficits on spatial working memory but not on verbal working memory or declarative memory. *J Sleep Res* 30: e13225, 2021. doi:10.1111/jsr.13225.
- Borland RG, Nicholson AN. Visual motor co-ordination and dynamic visual acuity. *Br J Clin Pharmacol* 18, Suppl 1: 69s–72s, 1984. doi:10.1111/j.1365-2125.1984.tb02583.x.
- Hart EP, Alvarez-Jimenez R, Davidse E, Doll RJ, Cohen AF, Van Gerven J, Groeneveld GJ. A computerized test battery to study

- pharmacodynamic effects on the central nervous system of cholinergic drugs in early phase drug development. *J Vis Exp* 144: e56569, 2019. doi:10.3791/56569.
22. **Kregenow DA, Swenson ER.** The lung and carbon dioxide: implications for permissive and therapeutic hypercapnia. *Eur Respir J* 20: 6–11, 2002. doi:10.1183/09031936.02.00400802.
23. **Laghi F, Shaikh H, Caccani N.** Basing intubation of acutely hypoxic patients on physiologic principles. *Ann Intensive Care* 14: 86, 2024. doi:10.1186/s13613-024-01327-w.
24. **Langford NJ.** Carbon dioxide poisoning. *Toxicol Rev* 24: 229–235, 2005. doi:10.2165/00139709-200524040-00003.
25. **Schulte JH.** Sealed environments in relation to health and disease. *Arch Environ Health* 8: 438–452, 1964. doi:10.1080/00039896.1964.10663693.
26. **National Archives.** Code of Federal Regulations. Ventilation, 14 CFR § 25.831 (Online). National Archives, 2026. <https://www.govinfo.gov/content/pkg/FR-1996-12-02/html/96-30525.htm>.
27. **Herbig B, Norrefeldt V, Mayer F, Reichherzer A, Lei F, Wargocki P.** Effects of increased recirculation air rate and aircraft cabin occupancy on passengers' health and well-being – results from a randomized controlled trial. *Environ Res* 216: 114770, 2023. doi:10.1016/j.envres.2022.114770.
28. **Harrison V, Mackenzie Ross SJ.** An emerging concern: toxic fumes in airplane cabins. *Cortex* 74: 297–302, 2016. doi:10.1016/j.cortex.2015.11.014.
29. **Li Y, Cheng P, Jia W.** Poor ventilation worsens short-range airborne transmission of respiratory infection. *Indoor Air* 32: e12946, 2022. doi:10.1111/ina.12946.
30. **Cadnum JL, Alhmidhi H, Donskey CJ.** Planes, trains, and automobiles: use of carbon dioxide monitoring to assess ventilation during travel. *Pathog Immun* 7: 31–40, 2022. doi:10.20411/pai.v7i1.495.
31. **Rosenberger W.** Effect of charcoal equipped HEPA filters on cabin air quality in aircraft. A case study including smell event related in-flight measurements. *Build Environ* 143: 358–365, 2018. doi:10.1016/j.buildenv.2018.07.031.
32. **Harvey TC, Raichle ME, Winterborn MH, Jensen J, Lassen NA, Richardson NV, Bradwell AR.** Effect of carbon dioxide in acute mountain sickness: a rediscovery. *Lancet* 2: 639–641, 1988 [Erratum in *Lancet* 2: 808, 1988]. doi:10.1016/s0140-6736(88)90465-5.
33. **Bärtsch P, Baumgartner RW, Waber U, Maggiorini M, Oelz O.** Comparison of carbon-dioxide-enriched, oxygen-enriched, and normal air in treatment of acute mountain sickness. *Lancet* 336: 772–775, 1990. doi:10.1016/0140-6736(90)93240-p.
34. **Van Dorp E, Los M, Dirven P, Sarton E, Valk P, Teppema L, Stienstra R, Dahan A.** Inspired carbon dioxide during hypoxia: effects on task performance and cerebral oxygen saturation. *Aviat Space Environ Med* 78: 666–672, 2007.
35. **Mutch WAC, Patel SR, Shahidi AM, Kulasekara SI, Fisher JA, Duffin J, Hudson C.** Cerebral oxygen saturation: graded response to carbon dioxide with isoxia and graded response to oxygen with isocapnia. *PLoS One* 8: e57881, 2013. doi:10.1371/journal.pone.0057881.
36. **Post TE, Heijn LG, Jordan J, van Gerven JMA.** Sensitivity of cognitive function tests to acute hypoxia in healthy subjects: a systematic literature review. *Front Physiol* 14: 1244279, 2023. doi:10.3389/fphys.2023.1244279.
37. **Martin-Gill C, Doyle TJ, Yealy DM.** In-flight medical emergencies: a review. *JAMA* 320: 2580–2590, 2018. doi:10.1001/jama.2018.19842.
38. **Coker RK, Armstrong A, Church AC, Holmes S, Naylor J, Pike K, Saunders P, Spurling KJ, Vaughn P.** BTS Clinical Statement on air travel for passengers with respiratory disease. *Thorax* 77: 329–350, 2022. doi:10.1136/thoraxjnl-2021-218110.
39. **Hammadah M, Kindya BR, Allard-Ratick MP, Jazbeh S, Eapen D, Wilson Tang WH, Sperling L.** Navigating air travel and cardiovascular concerns: is the sky the limit? *Clin Cardiol* 40: 660–666, 2017. doi:10.1002/clc.22741.
40. **Wan ZH, Wen FJ, Hu K.** Dynamic CO₂ inhalation: a novel treatment for CSR-CSA associated with CHF. *Sleep Breath* 17: 487–493, 2013. doi:10.1007/s11325-012-0719-x.